

ANDREW GOBEA:

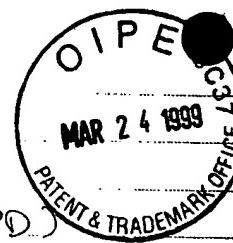


EXHIBIT
1

BBMM

RECEIVED 60CC COLD BLOOD (w CPD)
PCT mons 30%
T cells 70%

added 15 ml 1xPBS to bring to incubation
volume of 45ml.

Added 1.5 ml antibody (12.8)
incubated 25ml.

Formed Cellpro "ceprate".

Spin cells. Resuspended in PBS to
a final volume of 300cc in bag.

Ran through column.

Unabsorbed portion → spun down.
and consolidated in 1xPBS
for incubation.

75ml for incubation (added HPA)

1.5ml antibody (12.8) → spun. Spin down
following incubation. Resp to Vd. of 300cc in bag
Ran through 2nd column.

Skin cell portions from Runs 1 & 2
were combined (after counts, and
samples removed for staining)

Total cells 2.8×10^6 c for transduction

1L3 SANDO
WAN
1L6 SANDO
WAN
SLF AUG64
WAN

Final concen
will be dil
Cells are
concentrate

BBMM : FB
PBS

2.8×10^6 cells
PDT M

BMM + 31b1SCF (for 500ml of media)

D)

IL3 SANDOZ * 10230392 stock at 150ug/ml

want final: 20ng/ml $\times 2 \therefore 20\mu g$

133ul add

IL6 SANDOZ * 10450392 stock at 150ug/ml

want final: 50ng/ml $\times 2 \therefore 50\mu g$

333ul add

SLF AMBION * 1509F2 stock at 1.5mg/ml = 1500ug/ml

want final: 100ng/ml $\times 2 \therefore 100\mu g$

67ul add

Final concentrations are doubled since the media will be diluted 1:2 w/ viral supernatant. Cells are therefore incubated with the correct concentrations.

BMM: FBS Genuni lot# 10003H

BSA #115

2.8×10^6 cells want final: 5×10^4 cells

Put in 2 T-75 30ml each: 15ml B36S 051193

15ml LASN^G lot# 53

+ protamine sulfate 20ml
of 1:10 diluted stock

spin down
of 30ml in bag.

1/2
use
and

Cord Blood cells pre processing:

CFUs:

Set 143

Start:

Plate #	Sample	# Cells	# uL/ml media
- G418	1ab	5×10^4	50
+ G418	2ab		50
- G418	3ab		100
+ G418	4ab	1×10^5	100

absorb fraction:

absorb fraction

CFUs Post transduction: Set 144

Plate #	# Cells	# uL
- G418	500	7
1ab	1000	14
2ab	2000	28
3ab	500	7
+ G418	1000	14
4ab	2000	28
5ab		
6ab		

(yields)
absorb

COUNT:

$$\bar{x} = 34$$

$$\times 2 \times 10^4 = 6.8 \times 10^5 \text{ CFU}$$

$$\times 5.5 \text{ uL} = 3.7 \times 10^{10} \text{ C}$$

Reinfused on 5/15/93
Nontransduced T cell

G418

1ab	1000	20
2ab	2000	40

absorb
yields from
transduc.

Start:

$$5 \times 10^8 \text{ c}$$

PRE

$$0.71\%$$

Post ab

$$0.22\%$$

cell/ml media

$$\# 34t = 3.6 \times 10^6 \text{ c} = 1.1 \times 10^6 \text{ c}$$

30
20
100
100

absorbed fraction #1:

$$2 \times 10^6 \text{ c}$$

FL1 FL2 gate

$$31.94\%$$

FL1 FL2 gate

$$20.81\%$$

$$\# 34t = 0.64 \times 10^6 \text{ c}$$

$$= 0.42 \times 10^6 \text{ c}$$

absorbed fraction #2:

$$0.8 \times 10^6 \text{ c} \quad 2.41\%$$

$$5.80\%$$

$$\# 34t + c = 0.02 \times 10^6 \text{ c}$$

$$0.05 \times 10^6 \text{ c}$$

set 144

(yields)

absorbed #1:

PRE & FL1/FL2 gate

$$\frac{0.64 \times 10^6 \text{ c}}{3.6 \times 10^6 \text{ c}} = 17.8\%$$

PRE & FL1/FL2 gate

$$\frac{0.42 \times 10^6 \text{ c}}{3.6 \times 10^6 \text{ c}} = 11.7\%$$

post ab & FL1/FL2 gate

$$\frac{0.64 \times 10^6 \text{ c}}{1.1 \times 10^6 \text{ c}} = 58.2\%$$

Post ab & FL1/FL2 gate

$$\frac{0.42 \times 10^6 \text{ c}}{1.1 \times 10^6 \text{ c}} = 38.2\%$$

absorbed #2:

PRE & FL1/FL2

yields from
exclusion

exclusion

PRE & FL1/FL2

ZACHARY RIGGINS:

5/14/93

RECD 200cc COLD BLOOD

 inc
 12.8
 300
 PTT
 Ran

 PRE: mono 109 poly 109
 ✓

CVR

1

0

3x

3.

=

$$218 \times 50 = 10.9 \times 10^6 \text{ chnl}$$

$$\times 200\text{ml} = 2.2 \times 10^9 \text{ C}$$

start

 Added 3 vials (4.5ml) 12.8 ab
 inc. 25 min.

 spin down Rspd in 1xPBS to 300ml
 in bag.
per
fracce

com

26x1

wan

-2 =

spin down unabsorbed fraction for 2nd ab
incubation.spin stem cell fraction to Rspd in
smaller volume for count

13.51

CONT'S:

unabsorbed	
mono	poly
107	102
$109 \times 50 \times 10^3$	
$= 8.5 \times 10^6 \text{ chnl} \times 225\text{ml}$	
$= 1.9 \times 10^9 \text{ C}$	

stem	
mono	poly
172	16
$188 \times 2 \times 10^4$	
$= 3.8 \times 10^6 \text{ chnl} \times 55\text{ml}$	
$= 20.7 \times 10^6 \text{ C}$	

LAST

Incubated unadsorbed fraction w/ 4.5 ml
 12.8 ab. for 25 min.
 spun down
 Put in 300ml in bag (w/ 1x PBS)
 Ran through 2nd column.

Counts:

	<u>unadsorbed</u>		<u>stem</u>	
	monos	polys	monos	polys
start	30	33	58	4
	$6.3 \times 5.2 \times 10^3$		$6.2 \times 2 \times 10^4$	
	$3.15 \times 10^4 \text{ chnl} \times 600 \text{ ml}$		$= 1.2 \times 10^6 \text{ chnl}$	
	$= 1.9 \times 10^{10} \text{ C}$		$\times 5 \text{ ml} = 6 \times 10^6 \text{ C}$	
	↓			

20ml
 Percoll/Ficoll
 freeze → LWT(2)

combined stem cell fractions

26x10⁶ C for transduction
 want final [] = $5 \times 10^4 \text{ chnl}$

500 ml total

- 2 = 200ml supe
 200 ml media

13 flasks 40ml/flask 30ml supe
 20ml media (B36S)
 + 300ml protamine sulfate

L ASN supe 539 (bottles 18/19)

2×10^4
 ml $\times 5.5 \text{ ml}$

CFUs:

5/17/45

5/17

PRE

Plate #

Sample

Cells

cell

1ab

2ab

(-G418)

PRE + trans

↓ (+G418)

 5×10^4

↓

5

5

clear

Det

Recr

Post

BBMM + 316150F:

113 Sandoz * y0230292

OGI

116 Sandoz * y0450392

+G4

SCF AM(GEN) * 150952

↓

Took sample to micro for sterility ✓
each day of transduction pd.

OGI

+G4

Stat Gram stain done (negative)
before cells were given to baby.

5/15/93 4pm 2nd transduction:

Spun cells down from each flask

Rspd in fresh media & LASN supo

added Prostaglandin sulfate

5/16/93 3rd transduction 330pm

Repeated above.

5/17/93 cells washed 4x
 3x in 1x PBS + P15
 last wash in RPMI (no P15)

cells
5
5
count: $100 \times 10^6 \text{ c}$

$$\bar{x} = 15 \times 10 \times 10^4 = 15 \times 10^5 \text{ c}$$

$$\times 40 \text{ ml} = 100 \times 10^6 \text{ c}$$

Put in 5cc into 10cc syringe

Reinfused on 5/17/93 (UCSF)

Post trans. cells: 500/46

sample plate #	# cells	# el
DG418 1ab	500	4
↓ 2ab	1000	8
+G418 3ab	2000	16
↓ 4ab	500	4
+G418 5ab	1000	8
↓ 6ab	2000	16
DG418 7ab	1000	24
+G418 8ab	1000	24

my ✓

(time)

babey

iron:

n facsk

TSN supe

330PM